

A VERSATILE AND NEW HIGHLY STEREOSELECTIVE APPROACH TO THE SYNTHESIS
 OF L-GLYCERO-D-MANNO-HEPTOPYRANOSIDES

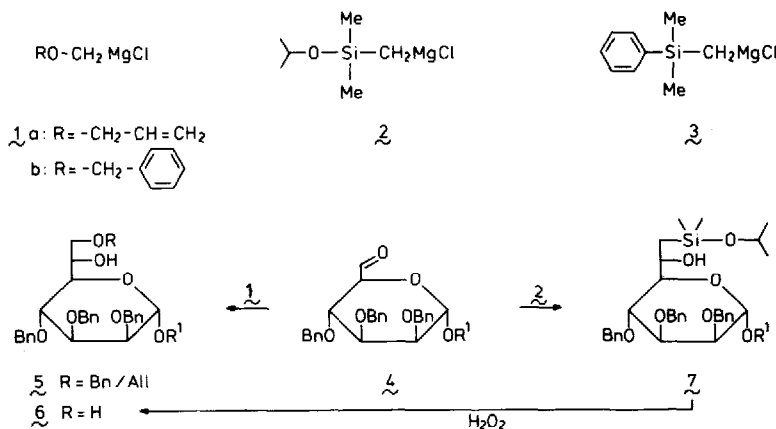
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Abstract: The reaction of (phenyldimethyl)silylmethylmagnesium chloride with benzyl 2,3,4-tri-O-benzyl- α -D-manno-hexodialdo-1,5-pyranoside affords the corresponding L-glycero-D-manno-addition product having at C-7 the phenyldimethylsilyl (PDMSi) group. The latter silanyl compound survived benzylation as well as glycosylation at OH-6, and the PDMSi group could be unmasked with overall retention of configuration to give a free hydroxyl.

It is well-established now that L-glycero-D-manno-heptopyranose (L-D-Hepp) is an important component of the core region of many polysaccharides of gram-negative bacteria¹. It has also been proposed that the core region can induce immunological reactions².

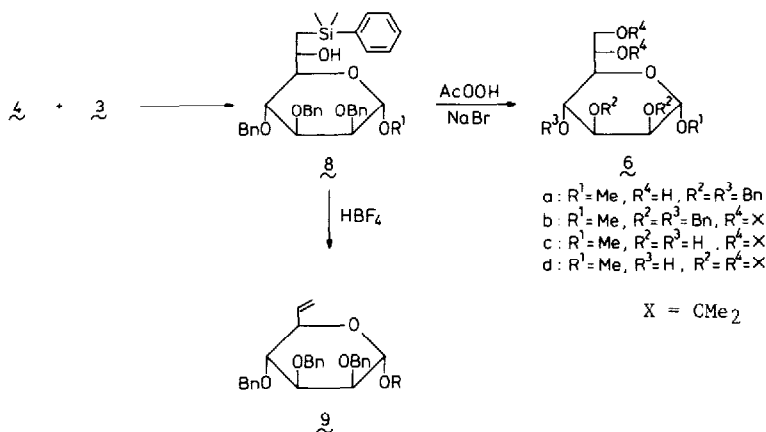
In a preliminary paper³ we reported that the benzyl-protected L-glycero-D-manno-heptose derivative **6** (R=H, R¹=Bn) could be prepared via the two-step Tamao procedure⁴ by reaction of benzyl 2,3,4-tri-O-benzyl- α -D-manno-hexodialdo-1,5-pyranoside (**4**) with (isopropoxydimethylsilyl)methylmagnesium chloride **2** followed by oxidation of the intermediate silane **7** with hydrogen peroxide. It was also pointed out that the above mentioned hydroxymethylation procedure was more reliable than the earlier proposed one-step method of Dziewiszek *et al.*⁵ which consisted of a Grignard reaction of the rather unstable⁶ allyl- or benzyloxymethylmagnesium chloride (**1**) with **4** to give **5**. However, the general usefulness of the Tamao hydroxymethylene extension method is, despite the high diastereoselective outcome of the reaction, not completely satisfactory to meet the demands for a wide variety of partially and suitably protected L-glycero-D-manno-heptopyranose derivatives.



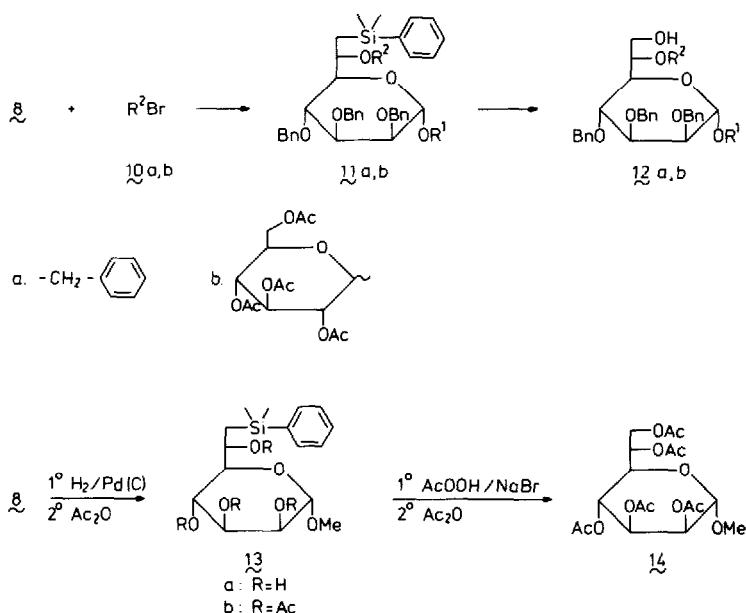
As part of a programme to study in detail the immunological reactions of the L-D-Hepp containing core region⁷ of *Neisseria meningitides*, we report, for the first time, that (phenyldimethylsilyl)methylmagnesium chloride (**3**) is a very convenient reagent for the syn-

thesis of partially protected L-glycero-D-manno-heptopyranosides (L-D-Hepp) in which the phenyldimethylsilyl at C-7 (e.g. 8) may function as a masked form of a hydroxyl group.

The reaction of 4 ($R^1=Bn$), with reagent 2 gives 7 containing the intrinsically labile isopropoxydimethylsilyl group which, for this reason, has to be desilylated *in situ* with hydrogen peroxide resulting in 6 having a free diol system. On the basis of earlier studies by Fleming *et al.*⁸ on silafunctional compounds, we reasoned that the corresponding more stable phenyldimethylsilyl (PDMSi) group could serve as a masked form of the hydroxy group. Thus reaction of fully-protected 4 with 3 would give 8 the C-6 hydroxy group of which will then be amenable to further processing (*i.e.* protection or interglycosydic bond formation to give 11a and 11b, respectively). Unmasking of the PDMSi group from 11a,b thus obtained by the recently developed two-step one-pot Fleming method^{8c} would result in the corresponding L-D-Hepp derivatives 12a,b.



The feasibility of the above mentioned approach was elaborated as follows. The aldehyde 4 ($R^1=Me$, 30 mmol, THF, 45 ml) was added to a cooled (0°C) solution of the easily accessible⁹ Grignard reagent 3 (60 mmol, THF, 45 ml). Work-up, after 1.5 h at 0°C , and purification (Silica gel chromatography) afforded chiral pure 8¹⁰ ($R^1=Me$, $[\alpha]_D^{20} + 11.5^\circ$, c 1, CHCl_3) in a yield of 70%. The sense of the diastereoselectivity was the same (*i.e.* predominant formation¹¹ of the L-D-Hepp derivative 8, $R^1=Me$), as corroborated earlier³ for 7 obtained by the reaction of 4 ($R^1=Bn$) with 2. Unmasking of the silyl group from 8 ($R^1=Me$) was easily accomplished^{8c} with $\text{AcOOH}/\text{NaBr}/\text{NaOAc}$ to furnish homogeneous¹² 6a in a yield of 85%. In this respect it is of interest to note that acidolysis of 8 ($R^1=Me$) in CH_2Cl_2 with tetrafluoroboric acid resulted in the quantitative formation of the Peterson¹³ elimination product 9 ($R^1=Me$, $[\alpha]_D^{20} + 24.6^\circ$, c 1; CHCl_3). The stability of the PDMSi group was further demonstrated by the preparation of the benzylated and glycosylated L-D-Hepp derivatives 11a and 11b. Thus benzylation of 8 ($R^1=Me$, 1 mmol) in DMF with sodium hydride (1.1 mmol) and benzyl bromide (10a, 1.2 mmol) in the presence of a catalytic amount of tetrabutylammonium iodide¹⁴ gave, after 1 h at 20°C and purification, homogeneous 11a ($[\alpha]_D^{20} + 24.6^\circ$, c 1, CHCl_3) in 95% yield. No trace of the elimination¹³ product 9 ($R^1=Me$) was formed under these mild benzylation conditions. Further, Koenigs-Knorr ($\text{HgCN}_2/\text{HgBr}_2$)-mediated glycosylation of 8 ($R^1=Me$, 1 mmol) in acetonitrile (10 ml) with the α -glucosyl bromide 10b (1.5 mmol)



resulted in the isolation of the $\beta(1-6)$ -linked disaccharide 11b ($\text{R}^1=\text{Me}$, $[\alpha]_{\text{D}}^{20} + 17.1^\circ$, c 1, CHCl_3) in a yield of 60%. Unmasking of the PDMSi from 11a and 11b obtained above, under the conditions mentioned before, afforded the corresponding derivatives 12a ($[\alpha]_{\text{D}}^{20} + 37.8^\circ$, c 1, CHCl_3) and 12b ($[\alpha]_{\text{D}}^{20} + 99.3^\circ$, c 1, CHCl_3), respectively, in a yield of 70%. Briefly summarized, the approach illustrated in this paper gives an easy access to L-D-Hepp building blocks, which may be glycosylated at C-6 (i.e., compound 8, $\text{R}^1=\text{Me}$) or C-7 (i.e. compound 12a, $\text{R}^1=\text{Me}$). On the other hand, compound 6a could be converted in an overall high yield by the following sequence of well-established reactions, i.e. acetonation (6a to 6b), hydrogenolysis (6b to 6c) and subsequent acetonation into the L-D-Hepp derivative 6d ($[\alpha]_{\text{D}}^{20} + 13.6^\circ$, c 1, CHCl_3) having a free C-4 hydroxy group. Finally, the stability of the PDMSi-group was further demonstrated by the smooth hydrogenolysis^{8b} ($\text{Pd(C)}/\text{H}_2$) of 8 ($\text{R}^1=\text{Me}$) resulting into 13a, which was then acetylated to give 13b ($[\alpha]_{\text{D}}^{20} + 23.5^\circ$, c 1, CHCl_3) in an excellent yield. The identity of 13b thus obtained was further corroborated by unmasking the PDMSi-function and subsequently acetylation to furnish 14 ($[\alpha]_{\text{D}}^{20} + 19.8^\circ$, c 1, CHCl_3) in an overall yield of 60%. At present we are studying in detail whether the PDMSi function in the non-acetylated product 13a survives acid-catalyzed acetalisation conditions which are required for the preparation of other protected L-D-Hepp derivatives.

In conclusion, the synthetic protocol described in this paper promises¹⁵ to be of great value for the preparation of antigenic oligosaccharides containing L-D-Hepp sugar units.

REFERENCES AND NOTES

- L. Kenne and B. Lindberg in *The Polysaccharides*, G.D. Aspinall (Ed.), vol. 2, pp. 287-363, Academic Press, New York, 1983. H. Brade and E.H. Rietschel, *Eur. J. Biochem.*, **145**, 231 (1984). H. Brade, U. Zahringner, E.H. Rietschel, R. Christian, G. Schulz and F.M. Unger, *Carbohydr. Res.*, **134**, 157 (1984). W. Kaca, J. de Jongh-Leuvenink, U.

- Zähringer, E.T. Rietschel, H. Brade, J. Verhoef, and V. Sinnwell, *Carbohydr. Res.*, **175**, 285 (1988).
2. H. Brade and C. Galopes, *Infect. Immun.*, **42**, 250 (1983).
 3. G.J.P.H. Boons, F.A.M. van der Klein, G.A. van der Marel and J.H. van Boom, *Recl. Trav. Chim. Pays-Bas*, **107**, 507 (1988).
 4. K. Tamao and N. Yshida, *Tetrahedron Lett.*, **25**, 4245 (1984).
 5. K. Dziewiszek and A. Zamojski, *Carbohydr. Res.*, **150**, 163 (1986).
 6. B. Castro, *Bull. Soc. Chim. France*, 1533 (1967).
 7. H.J. Jennings, M. Beurret, A. Gamian, F. Michon, *Antonie van Leeuwenhoek*, **53**, 519 (1987).
 8. a) W. Bernhard, I. Fleming and D. Waterson, *J. Chem. Soc., Chem. Commun.* 28 (1984).
 b) I. Fleming, R. Henning and H. Plaut, *ibid*, 29 (1984).
 c) I. Fleming and Philip E.J. Sanderson, *Tetrahedron Lett.*, **28**, 4229 (1987).
 9. L.H. Sommer, I.R. Gold, G.M. Goldberg and A.S. Marans, *J. Am. Chem. Soc.*, **71**, 1509 (1949).
 10. Satisfactory elemental analytical data were obtained for compounds 6d, 8, 9, 11a,b, 12a,b, 13b and 14.
 Relevant ¹H-NMR (CDCl₃) data of compound: 6d; δ 4.94 (s, 1H, H₁), 4.39 (dt, 1H, H₆, J_{5,6} = 5.0 Hz, J_{6,7} = 7.0 Hz), 4.07 (dd, 1H, H₇, J_{7,7'} = 8.5 Hz), 3.96 (dd, 1H, H_{7'}). 8; δ 4.71 (d, 1H, H₁, J_{1,2} = 2.0 Hz), 1.37 (dd, 1H, H₇, J_{7,7'} = 17.0 Hz, J_{6,7} = 12.0 Hz), 0.94 (dd, 1H, J_{7,7'}, J_{6,7} = 5.0 Hz) 0.36, 0.35 (2xS, 6H, Si(CH₃)₂). 9; δ 6.02 (M, 1H, H₆), 5.37 (M, 2H, H₇, H_{7'}), 4.71 (d, 1H, H₁, J_{1,2} = 2.0 Hz). 11a; δ 4.85 (d, 1H, H₁, J_{1,2} = 2.0 Hz), 1.42 (m, 2H, H₇, H_{7'}), 0.33 (s, 6H, Si(CH₃)₂). 11b; δ 4.93 (d, 1H, H₁), 4.79 (d, 1H, H₁, J_{1,2} = 2.0 Hz), 1.55 (M, 2H, H₇, H_{7'}). 12a; δ 4.84 (d, 1H, H₁, J_{1,2} = 2.0 Hz, 3.98 - 3.90 (m, 4H, H₃, H₆, H₇, H_{7'}). 14; δ 4.74 (d, 1H, H₁, J_{1,2} = 1.5 Hz), 4.36 - 4.21 (m, 2H, H₇, H_{7'}), 1.92 - 2.21 (5xs, 15H, 5xCH₃Ac).
 Relevant ¹³C-NMR (CDCl₃) data of compound: 6d; δ 98.37 (C₁), 65.58 (C₇). 8; δ 99.39 (C₁), 21.72 (C₇), -2.45 (Si(CH₃)₂). 9; δ 135.43 (C₆), 117.88 (C₇), 99.04 (C₁). 11a; δ 99.08 (C₁), 17.02 (C₇), -2.13, -2.33 (Si(CH₃)₂). 11b; δ 101.91 (C_{1'}), 98.64 (C₁), 20.03 (C₇), -2.21 (Si(CH₃)₂). 12a; δ 98.84 (C₁), 62.05 (C₇). 12b; δ 101.50 (C_{1'}), 98.35 (C₁), 64.18, 61.70 (C_{6'}, C₇). 13b; δ 98.93 (C₁), 19.80 (C₇), 1.05, 0.76 (Si(CH₃)₂). 14; δ 98.93 (C₁), 61.67 (C₇).
 11. In this particular case the formation of the D-D-Hepp stereoisomer was negligible.
 12. Conversion of 8 into 6a proceeded as expected (ref. 8c) with retention of configuration. The latter was confirmed by hydrogenolysis followed by conversion of the de-benzylated product into the known diethylthioacetal (see ref. 15a).
 13. D. Peterson, *J. Org. Chem.*, **33**, 780 (1968).
 14. S. Czernecki, G. Georgoulis and C. Provelenghion, *Tetrahedron Lett.*, **17**, 3535 (1976).
 15. Apart from the synthesis of L-D-Hepp derivatives by Dziewiszek *et al.* (ref. 5), two other rather time-consuming approaches have been published. See: (a) H. Teuber *et al.*, *Biochemistry*, **7**, 3303 (1968), and (b) H. Paulsen *et al.*, *Liebigs Ann. Chem.*, 675 (1986).

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